



Recent Research in Hurler Disease (MPS I) in Childhood

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Letter to the Editor

Hurler syndrome is the most severe form of the rare metabolic disorder mucopolysaccharidosis type 1, caused by an enzyme deficiency that prevents the breakdown of certain sugars, leading to the accumulation of dermatansulfate and heparansulfate in tissues [1,2]. The gene defect is located at chromosome 4p16.3 gene locus. The incidence is 1:145.000 respectively. It is a monogenetic disease with an autosomal recessive trait. The gene defect leads to an enzyme defect of the enzyme alpha-L-iduronidase (IDUA) with the result of extensive accumulation of glycosaminoglycans in different cell tissues [3,4]. Newborn screening detection is crucial for therapy success, especially for neuroprotection [3]. There are 3 different forms of Hurler disease. Morbus Hurler shows a serious involvement of the CNS, Morbus Hurler-Scheie an attenuated form with an incidence of 1:280000, and Morbus Scheie, which is found in 1:500000 newborns, shows only a moderate form of the disease [5-8]. Typical symptoms include coarse facial features, skeletal changes, enlarged organs with hepatosplenomegaly, heart and breathing problems, and intellectual disabilities due to central nervous system involvement [9,10]. Early diagnosis and specialized therapies such as stem cell transplantation are important, but the prognosis is often limited due to the severity of the condition [11,12]. The challenge treating the disease in childhood is delivering sufficient enzymes to the brain to ameliorate the cognitive brain development by accumulation of GAG in the brain cells [13-15]. Recent research is focusing on next generation enzyme replacement therapies with crossing the blood-brain-barrier, HSCT, targeting nonsense mutations at mRNA level through nonsense suppression

and mRNA editing suppressor t^rNA's, in vivo and ex vivo gene transfers [16-20].

In Europe, the prevalence of the Hurler subtype of MPS1 is estimated at 1/200,000. The main features of Hurler syndrome are skeletal deformities and delayed motor and intellectual development. Patients typically present in the first year of life with changes in the musculoskeletal system: short stature, dysostosis multiplex, thoracolumbar kyphosis, progressive coarsening of facial features, large head, frontal bossing, flat nasal bridge with broad nasal tip and anteverted nares, full cheeks, widened lips, cardiomyopathy with valve abnormalities, sensorineural hearing loss, enlarged tonsils and adenoids, nasal discharge. Developmental delay becomes apparent between 12 and 24 months, especially in the area of speech, followed by progressive cognitive and sensory decline. Hydrocephalus may develop in some patients by age 2 [21]. By age 3, diffuse corneal changes progress to corneal opacification. Organomegaly, hernias, and hirsutism are additional symptoms. Hurler syndrome is caused by mutations in the IDUA gene, leading to complete loss of alpha-L-iduronidase activity and lysosomal storage of Dermatan Sulfate (DS) and Heparan Sulfate (HS). Hurler syndrome is inherited in an autosomal recessive manner. Early diagnosis is challenging as the initial clinical signs are nonspecific. However, early diagnosis is crucial for initiating treatment as soon as possible. Laboratory diagnosis involves detecting increased excretion of HS and DS in urine using the DMB test and GAG electrophoresis, as well as demonstrating enzyme deficiency in leukocytes or fibroblasts. Molecular testing is available. Differential

diagnoses include the milder form of mucopolysaccharidosis type 1, Hurler-Scheie syndrome, although this form is associated with only mild cognitive impairment. Differential diagnoses also include mucopolysaccharidosis type 6 and type 2, and mucopolysaccharidosis type 2. Prenatal diagnosis is possible enzymatically or molecularly. Genetic counseling should be offered to affected families. Management is multidisciplinary. Hematopoietic Stem Cell Transplantation (HSCT) is the treatment of choice for patients with Hurler syndrome under 2.5 years (and selected patients over this age) as it can prolong survival, preserve neurocognition, and improve some somatic features. HSCT should be performed early in the disease course before developmental decline begins. Enzyme replacement therapy with laronidase is recommended for all Hurler patients and is a lifelong therapy that alleviates non-neurological symptoms. Early ERT has been shown to delay or even prevent the development of some clinical features of the disease. Additional management of Hurler syndrome is largely supportive and includes surgical interventions (e.g., adenotonsillectomy, hernia repair, ventriculoperitoneal shunt, heart valve replacement, carpal tunnel release, spinal decompression), physical therapy, occupational therapy, speech therapy, respiratory support like continuous positive airway pressure with oxygen supplementation, hearing aids, and medications for pain and gastrointestinal disturbances. Patients often succumb to the disease in the first decade due to respiratory and cardiac complications, but ERT and HSCT can improve life expectancy. The timing of diagnosis and initiation of treatment is a crucial factor for the success of HSCT and laronidase.

Until the age of about 2.5 years, bone marrow or hematopoietic stem cell transplantation is the treatment of choice for patients with M. Hurler. Through the transplantation of bone marrow from a suitable donor, the patient receives blood cells that can produce the enzyme alpha-L-iduronidase. These cells release some of the produced, intact enzyme into the environment, which can be taken up by other body cells and transported into their lysosomes. This allows the stored glycosaminoglycans to be broken down. However, transplantation can only positively influence cognitive development and life expectancy. HSCT cannot cure the disease. In patients with M. Hurler, enzyme replacement therapy is often used before or after bone marrow or hematopoietic stem cell transplantation. In this therapy, the defective enzyme is replaced by a biotechnologically produced form of the human enzyme, allowing the pathological storage of glycosaminoglycans to be broken down again. However, due to the blood-brain barrier, the enzyme replacement therapy does not reach the central nervous system, so this therapy cannot directly affect the cognitive and motor symptoms of M. Hurler. This can currently only be achieved through early stem cell transplantation. Before HSCT, enzyme replacement therapy can improve the overall condition of patients. Additionally, enzyme replacement therapy can support the transplantation and alleviate symptoms, as MPS I cannot be cured by HSCT. For M. Hurler patients who are diagnosed later than 2.5 years and for

whom bone marrow or stem cell transplantation is no longer an option, enzyme replacement therapy is available to treat the non-neurological manifestations of the disease.

The future prospects for Hurler syndrome are improving due to therapy-specific advances, with gene therapy and next generation enzyme replacement therapy as well as early hematopoietic stem cell transplants being the main pillars that increase life expectancy and alleviate symptoms, even though a complete cure is still pending [22]. The key lies in early diagnosis to protect brain and skeletal development, which are currently the biggest challenges [23-26]. New biomarkers are under research in animal models [27,28]. Enzyme replacement therapy is based on providing the missing substance through recombinant enzymes, non-neurological symptoms are improved and quality of life is increased, but it does not cross the blood-brain barrier to date [23,29-37]. Hematopoietic Stem Cell Transplantation (HSCT) is focused on younger patients under 2.5 years, it can preserve neurocognitive development and extend life expectancy, and is the most effective method so far, although it carries risks and requires a donor search [38-40]. Treats specific complaints such as heart or breathing problems and improves daily life.

Gene therapy aims to directly correct the gene defect and has the potential to address the root cause of the disease, enabling more comprehensive treatments [21,41-52]. Nanovector research is of utmost importance to deliver a functioning enzyme into brain cells with crossing the blood brain barrier [53-58]. Further research focus on intranasal delivery of an AAV-vector gene delivery system [59-61]. Substrate reduction therapy aims to reduce the production of harmful substances. The combination of ERT with gene therapy or SRT is being researched to increase effectiveness and also reach the brain [62]. The biggest challenge is the CNS involvement, therefore overcoming the blood-brain barrier remains the biggest hurdle for ERT.

In conclusion, with innovative approaches such as gene therapy with AAV-vector gene-or enzyme delivery systems or CRISP/Cas9 therapy and improved stem cell transplantation procedures, there is increasing hope to better control the life-threatening aspects of Hurler syndrome and improve the quality of life, even though the path to a complete cure is not yet complete [63-65].

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Conflict of Interest

None.

References

1. Clarke LA (2025) Mucopolysaccharidosis Type I. 2002 Oct 31 [updated 2025 Dec 4]. In: Adam MP, Bick S, Mirzaa GM, Pagon RA, Wallace SE, Amemiya A, editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle 1993-2026.

2. Machnikowska Sokołowska M, Myszczyk A, Wieszała E, Wieja Błach D, Jamroz E, et al. (2023) Mucopolysaccharidosis Type 1 among Children-Neuroradiological Perspective Based on Single Centre Experience and Literature Review. *Metabolites* 13(2): 209.
3. Fillman T, Matteson J, Tang H, Mathur D, Zahedi R, et al. (2023) First Three Years Experience of Mucopolysaccharidosis Type-I Newborn Screening in California. *J Pediatr* 263: 113644.
4. Liu D, Jiang Z, Deng L, Li H, Jiang H (2023) Identification of an α -L-iduronidase (IDUA) M1T mutation in a Chinese family with autosomal recessive mucopolysaccharidosis I. *Ann N Y Acad Sci* 1526(1): 114-125.
5. Ramarajan MG, Parthasarathy KTS, Gaikwad KB, Joshi N, Garapati K, et al. (2024) Alterations in Hurler-Scheie Syndrome Revealed by Mass Spectrometry-Based Proteomics and Phosphoproteomics Analysis *OMICS* 28(11): 548-562.
6. Lamichhane S, Sapkota A, Sapkota S, Adhikari N, Aryal S, et al. (2023) Mucopolysaccharidosis type I Hurler-Scheie syndrome: a case report. *Ann Med Surg (Lond)* 86(1): 588-593.
7. Ahmed A, Ou L, Rudser K, Shapiro E, Eisengart JB, et al. (2019) A longitudinal study of neurocognition and behavior in patients with Hurler-Scheie syndrome heterozygous for the L238Q mutation. *Mol Genet Metab Rep* 20: 100484.
8. Donati MA, Pasquini E, Spada M, Polo G, Burlina A (2018) Newborn screening in mucopolysaccharidoses. *Ital J Pediatr* 44: 126.
9. Grosse SD, Lam WKK, Wiggins LD, Kemper AR (2017) Cognitive outcomes and age of detection of severe mucopolysaccharidosis type I. *Genet Med* 19(9): 975-982.
10. Ou L, Przybilla MJ, Whitley CB (2017) Proteomic analysis of mucopolysaccharidosis I mouse brain with two-dimensional polyacrylamide gel electrophoresis. *Mol Genet Metab* 120(1-2): 101-110.
11. Pontesilli S, Baldoli C, Rosa PAD, Cattoni A, Bernardo ME, et al. (2022) Evidence of Treatment Benefits in Patients with Mucopolysaccharidosis Type I-Hurler in Long-term Follow-up Using a New Magnetic Resonance Imaging Scoring System. *J Pediatr* 240: 297-301.
12. Kingma SDK, Jonckheere AI (2021) MPS I: Early diagnosis, bone disease and treatment, where are we now? *J Inher Metab Dis* 44(6): 1289-1310.
13. Guffon N, Pettazzoni M, Pangaud N, Reynes N, Le Peillet Feuillet E, et al. (2025) Clinical outcomes of exclusive enzyme therapy (laronidase) in a cohort of patients with mucopolysaccharidosis type I. *Orphanet J Rare Dis* 21(1): 11
14. Toscano A, Musumeci O, Sacchini M, Ravaglia S, Siciliano G, et al. (2025) Long-term safety outcomes and patient preferences for home-based intravenous enzyme replacement therapy (ERT) in Pompe disease and Mucopolysaccharidosis Type I (MPS-I): final results of two-year observation. *Orphanet J Rare Dis* 20(1): 639.
15. Rabbani A, Alaei M, Asl SN, Setoodeh A, Shakiba M, et al. (2025) Efficacy and safety of a biosimilar laronidase versus the reference laronidase in patients with mucopolysaccharidosis type I. *Sci Rep* 15(1): 30427.
16. Pierce SE, Erwood S, Oye K, An M, Krasnow N, et al. (2025) Prime editing-installed suppressor tRNAs for disease-agnostic genome editing. *Nature* 648(8092): 191-202.
17. Tucci F, Uria Oficialdegui ML, Consiglieri G, Cossutta M, Filisetti C, et al. (2026) Non-neurological, non-skeletal outcomes after hematopoietic stem and progenitor cell-gene therapy (OTL-203) for Hurler syndrome. *Mol Ther* 34(1): 443-454.
18. Agranian M, Demurger F, Dubourg C, Fromageot J, Dufour AC, et al. (2025) Prenatal diagnosis of mucopolysaccharidosis type I on hepatosplenomegaly and coarse features: a case-report. *BMC Pregnancy Childbirth* 25(1): 3.
19. Harmatz P, Prada CE, Burton BK, Lau H, Kessler CM, et al. (2022) First-in-human in vivo genome editing via AAV-zinc-finger nucleases for mucopolysaccharidosis I/II and hemophilia B. *Mol Ther* 30(12): 3587-3600.
20. Schuh RS, Bidone J, Poletto E, Pinheiro CV, Pasqualim G, et al. (2018) Nasal Administration of Cationic Nanoemulsions as Nucleic Acids Delivery Systems Aiming at Mucopolysaccharidosis Type I Gene Therapy. *Pharm Res* 35(11): 221.
21. Huang S, Nascene DR, Shanley R, Choi M, Lund TC, et al. (2025) Longitudinal clinical and imaging analysis of hydrocephalus in a single-center study in 57 patients with mucopolysaccharidosis type IH (Hurler syndrome). *J Neurosurg Pediatr* 36(2): 157-164.
22. Horovitz DD, Acosta AX, Giugliani R, Hlavatá A, Hlavatá K, et al. (2016) Alternative laronidase dose regimen for patients with mucopolysaccharidosis I: a multinational, retrospective, chart review case series. *Orphanet J Rare Dis* 11(1): 51.
23. Kida S, Koshimura Y, Yoden E, Yoshioka A, Morimoto H, et al. (2023) Enzyme replacement with transferrin receptor-targeted α -L-iduronidase rescues brain pathology in mucopolysaccharidosis I mice. *Mol Ther Methods Clin Dev* 29: 439-449.
24. Bay L, Amartino H, Antacle A, Arberas C, Berretta A, et al. (2021) New recommendations for the care of patients with mucopolysaccharidosis type I. *Arch Argent Pediatr* 119(2): e121-e128.
25. Nan H, Park C, Maeng S (2020) Mucopolysaccharidoses I and II: Brief Review of Therapeutic Options and Supportive/Palliative Therapies. *Biomed Res Int* 2020: 2408402.
26. D'Avanzo F, Rigon L, Zanetti A, Tomanin R (2020) Mucopolysaccharidosis Type II: One Hundred Years of Research, Diagnosis, and Treatment. *Int J Mol Sci* 21(4): 1258.
27. Zhang C, Gawri R, Lau YK, Spruce LA, Fazelinia H, et al. (2023) Proteomics identifies novel biomarkers of synovial joint disease in a canine model of mucopolysaccharidosis I. *Mol Genet Metab* 138(2): 107371.
28. Langereis EJ, van Vlies N, Church HJ, Geskus RB, Hollak CE, (2015) Biomarker responses correlate with antibody status in mucopolysaccharidosis type I patients on long-term enzyme replacement therapy. *Mol Genet Metab* 114(2): 129-137.
29. Shinoda C, Kitakaze K, Sasai Y, Nishioka SI, Kobayashi I, et al. (2025) N-glycan-modified α -L-iduronidase produced by transgenic silkworms ameliorates clinical signs in a Japanese macaque with mucopolysaccharidosis I. *Commun Med (Lond)* 5(1): 128.
30. Rintz E, Ziemian M, Kobus B, Gaffke L, Pierzynowska K, et al. (2024) Synergistic effects of resveratrol and enzyme replacement therapy in the Mucopolysaccharidosis type I. *Biochem Pharmacol* 229: 116467.
31. Toscano A, Musumeci O, Sacchini M, Ravaglia S, Siciliano G, et al. (2023) Safety outcomes and patients' preferences for home-based intravenous enzyme replacement therapy (ERT) in pompe disease and mucopolysaccharidosis type I (MPS I) disorder: COVID-19 and beyond. *Orphanet J Rare Dis* 18(1): 338.
32. Zhu W, Ou L, Zhang L, Clark IH, Zhang Y, et al. (2023) Mapping brain networks in MPS I mice and their restoration following gene therapy. *Sci Rep* 13(1): 12716.
33. Lin Y, Wang X, Rose KP, Dai M, Han J, et al. (2020) miR-143 Regulates Lysosomal Enzyme Transport across the Blood-Brain Barrier and Transforms CNS Treatment for Mucopolysaccharidosis Type I. *Mol Ther* 28(10): 2161-2176.
34. Jameson E, Jones S, Remington T (2019) Enzyme replacement therapy with laronidase (Aldurazyme®) for treating mucopolysaccharidosis type I. *Cochrane Database Syst Rev* 6(6): CD009354.
35. Concolino D, Deodato F, Parini R (2018) Enzyme replacement therapy: efficacy and limitations. *Ital J Pediatr* 44(Suppl 2): 120.

36. Giugliani R, Giugliani L, de Oliveira Poswar F, Donis KC, Corte AD, et al. (2018) Neurocognitive and somatic stabilization in pediatric patients with severe Mucopolysaccharidosis Type I after 52 weeks of intravenous brain-penetrating insulin receptor antibody-iduronidase fusion protein (valanafusp alpha): an open label phase 1-2 trial. *Orphanet J Rare Dis* 13(1): 110.
37. Partridge WM, Boado RJ, Giugliani R, Schmidt M (2018) Plasma Pharmacokinetics of Valanafusp Alpha, a Human Insulin Receptor Antibody-Iduronidase Fusion Protein, in Patients with Mucopolysaccharidosis Type I. *BioDrugs* 32(2): 169-176.
38. Orchard PJ, Gupta AO, Eisengart JB, Polgreen LE, Pollard LM, et al. (2022) Hematopoietic stem cell transplant for Hurler syndrome: does using bone marrow or umbilical cord blood make a difference? *Blood Adv* 6(23): 6023-6027.
39. Sevilla J, Iriondo J, Sebastian E, Gonzalez-Vicent M, Schwartz JD, et al. (2022) Letter to the Editor:Hematopoietic Stem and Progenitor Cell Mobilization and Collection for Patients Diagnosed with Osteopetrosis and Hurler Syndrome. *Hum Gene Ther* 33 (3-4): 213-214.
40. Gentner B, Tucci F, Galimberti S, Fumagalli F, De Pellegrin M, et al. (2021) Hematopoietic Stem- and Progenitor-Cell Gene Therapy for Hurler Syndrome. *N Engl J Med* 385(21): 1929-1940.
41. Wood SR, Bigger BW (2022) Delivering gene therapy for mucopolysaccharide diseases. *Front Mol Biosci* 9: 965089.
42. Jin X, Su J, Zhao Q, Li R, Xiao J, et al. (2022) Liver-directed gene therapy corrects neurologic disease in a murine model of mucopolysaccharidosis type I-Hurler. *Mol Ther Methods Clin Dev* 25: 370-381.
43. Hurt SC, Dickson PI, Curiel DT (2021) Mucopolysaccharidoses type I gene therapy. *J Inher Metab Dis* 44(5): 1088-1098.
44. Ou L, Przybilla MJ, Ahlat O, Kim S, Overn P, et al. (2020) A Highly Efficacious PS Gene Editing System Corrects Metabolic and Neurological Complications of Mucopolysaccharidosis Type I. *Mol Ther* 28(6): 1442-1454.
45. Poletto E, Baldo G, Gomez-Ospina N (2020) Genome Editing for Mucopolysaccharidoses. *Int J Mol Sci* 21(2): 500.
46. Squeri G, Passerini L, Ferro F, Laudisa C, Tomasoni D, et al. (2019) Targeting a Pre-existing Anti-transgene T Cell Response for Effective Gene Therapy of MPS-I in the Mouse Model of the Disease. *Mol Ther* 27(7): 1215-1227.
47. Ou L, DeKolver RC, Rohde M, Tom S, Radeke R, et al. (2019) ZFN-Mediated In Vivo Genome Editing Corrects Murine Hurler Syndrome. *Mol Ther* 27(1): 178-187.
48. Schuh RS, Poletto E, Pasqualim G, Tavares AMV, Meyer FS, et al. (2018) In vivo genome editing of mucopolysaccharidosis I mice using the CRISPR/Cas9 system. *J Control Release* 288: 23-33.
49. Eisengart JB, Rudser KD, Xue Y, Orchard P, Miller W, et al. (2018) Long-term outcomes of systemic therapies for Hurler syndrome: an international multicenter comparison. *Genet Med* 20(11): 1423-1429.
50. Lau AA, Hemsley KM (2017) Adeno-associated viral gene therapy for mucopolysaccharidoses exhibiting neurodegeneration. *J Mol Med (Berl)* 95(10): 1043-1052.
51. Penati R, Fumagalli F, Calbi V, Bernardo ME, Aiuti A (2017) Gene therapy for lysosomal storage disorders: recent advances for metachromatic leukodystrophy and mucopolysaccharidosis I. *J Inher Metab Dis* 40(4): 543-554.
52. Brown N, Song L, Kollu NR, Hirsch ML (2017) Adeno-Associated Virus Vectors and Stem Cells: Friends or Foes? *Hum Gene Ther* 28(6): 450-463.
53. Da Ros T, Ostric A, Andreola F, Filocamo M, Pietrogrande M, et al. (2018) Carbon nanotubes as nanovectors for intracellular delivery of iduronidase in Mucopolysaccharidosis type I. *Nanoscale*. 10(2): 657-665.
54. Holley RJ, Ellison SM, Fil D, O'Leary C, McDermott J, et al. (2018) Bigger BW. Macrophage enzyme and reduced inflammation drive brain correction of mucopolysaccharidosis IIIB by stem cell gene therapy. *Brain* 141(1): 99-116.
55. Fraga M, de Carvalho TG, Bidone J, Schuh RS, Matte U, et al. (2017) Factors influencing transfection efficiency of pIDUA/nanoemulsion complexes in a mucopolysaccharidosis type I murine model. *Int J Nanomedicine* 12: 2061-2067.
56. Fraga M, de Carvalho TG, Diel Dda S, Kretzmann Filho NA, Teixeira HF, et al. (2015) Cationic Nanoemulsions as a Gene Delivery System: Proof of Concept in the Mucopolysaccharidosis I Murine Model. *J Nanosci Nanotechnol* 15(1): 810-6.
57. Sharma R, Anguela XM, Doyon Y, Wechsler T, DeKolver RC, et al. (2015) In vivo genome editing of the albumin locus as a platform for protein replacement therapy. *Blood* 126(15): 1777-84.
58. Fraga M, Bruxel F, Diel D, de Carvalho TG, Perez CA, et al. (2015) PEGylated cationic nanoemulsions can efficiently bind and transfect pIDUA in a mucopolysaccharidosis type I murine model. *J Control Release* 209: 37-46.
59. Belur LR, Temme A, Podetz-Pedersen KM, Riedl M, Vulchanova L, et al. (2017) Intranasal Adeno-Associated Virus Mediated Gene Delivery and Expression of Human Iduronidase in the Central Nervous System: A Noninvasive and Effective Approach for Prevention of Neurologic Disease in Mucopolysaccharidosis Type I. *Hum Gene Ther* 28(7): 576-587.
60. Ou L, Przybilla MJ, Koniar BL, Whitley CB (2016) Elements of lentiviral vector design toward gene therapy for treating mucopolysaccharidosis I. *Mol Genet Metab Rep* 8: 87-93.
61. Hinderer C, Bell P, Louboutin JP, Katz N, Zhu Y, et al. (2016) Neonatal tolerance induction enables accurate evaluation of gene therapy for MPS I in a canine model. *Mol Genet Metab* 119(1-2): 124-30.
62. Santi L, De Ponti G, Dina G, Pievani A, Corsi A, et al. (2020) Neonatal combination therapy improves some of the clinical manifestations in the Mucopolysaccharidosis type I murine model. *Mol Genet Metab* 130(3): 197-208.
63. Vera LNP, Schuh RS, Fachel FNS, Poletto E, Piovesan E, et al. (2022) Brain and visceral gene editing of mucopolysaccharidosis I mice by nasal delivery of the CRISPR/Cas9 system. *J Gene Med* 24(4): e3410.
64. Carneiro P, de Freitas MV, Matte U (2022) In silico analysis of potential off-target sites to gene editing for Mucopolysaccharidosis type I using the CRISPR/Cas9 system: Implications for population-specific treatments. *PLoS One* 17(1): e0262299.
65. Schuh RS, Gonzalez EA, Tavares AMV, Seolin BG, Elias LS, et al. (2020) Neonatal nonviral gene editing with the CRISPR/Cas9 system improves some cardiovascular, respiratory, and bone disease features of the mucopolysaccharidosis I phenotype in mice. *Gene Ther* 27(1-2): 74-84.